

OBSTETRICS

First-trimester metabolomic detection of late-onset preeclampsia

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OBJECTIVE: We sought to identify first-trimester maternal serum biomarkers for the prediction of late-onset preeclampsia (PE) using metabolomic analysis.

STUDY DESIGN: In a case-control study, nuclear magnetic resonance-based metabolomic analysis was performed on first-trimester maternal serum between 11⁺⁰-13⁺⁶ weeks of gestation. There were 30 cases of late-onset PE, ie, requiring delivery ≥ 37 weeks, and 59 unaffected controls. The concentrations of 40 metabolites were compared between the 2 groups. We also compared 30 early-onset cases to the late-onset group.

RESULTS: A total of 14 metabolites were significantly elevated and 3 significantly reduced in first-trimester serum of late-onset PE patients. A complex model consisting of multiple metabolites and maternal demographic characteristics had a 76.6% sensitivity at 100% specificity for PE detection. A simplified model using fewer predictors yielded 60% sensitivity at 96.6% specificity. Strong separation of late- vs early-onset PE groups was achieved.

CONCLUSION: Significant differences in the first-trimester metabolites were noted in women who went on to developed late-onset PE and between early- and late-onset PE.

Key words: metabolomics, preeclampsia prediction

Cite this article as: Bahado-Singh RO, Akolekar R, Mandal R, et al. First-trimester metabolomic detection of late-onset preeclampsia. *Am J Obstet Gynecol* 2013;208:58.e1-7.

Metabolomics, a relatively recent addition to the “omics” family, involves the high-throughput characterization and interpretation of the small-molecule metabolites (<1500 d) produced by cells, tissues, and organisms. To date, >8000 human metabolites from >80 chemical classes have been identified or catalogued.¹ As technology im-

proves it is expected that this number could grow by a factor of ≥ 10 .² Because of the wide chemical diversity of metabolites, their tight coupling with environmental interactions (food, drugs, gut microbiota), and their huge phenotypic-dependent concentration variations ($\geq 10^6$), metabolomics offers a powerful, quantitative route to describe the actual phenotype of cells, tissues, or organisms in both normal and diseased states. Recently, significant advances have occurred both in metabolite identification techniques^{1,2} and computational techniques³ for analyzing the large volume of data generated by metabolomic studies. There is currently tremendous interest in the use of metabolomics for the characterization and early diagnosis of complex diseases.⁴

Preeclampsia (PE) is a common obstetric disorder characterized by hypertension and proteinuria during pregnancy. It is a cause of significant morbidities, affecting the health of both the mother⁵ and fetus. However, its causes and pathophysiology largely remain a mystery. It now appears that PE is at least 2 fairly distinct disorders, an early-onset and a late-onset form.^{6,7} The early-onset variety typically occurs <34-35 weeks of pregnancy, and

is associated with significant fetal morbidities. The pathophysiology is thought to be failure of trophoblast invasion of the maternal spiral arteriole⁸ resulting in maintenance of high maternal vascular resistance. This is consistent with the high frequency of placental underperfusion reported⁹ in this disorder.

The late-onset form is considered to be more of a maternal constitutional disorder¹⁰ due to underlying maternal microvascular disorders such as hypertension or a genetic predisposition in which poor trophoblast invasion is thought to play a less significant role. Late-onset PE is significantly more common and while it often has a mild course can be associated with significant clinical morbidities.¹¹ It is therefore important to investigate its pathogenesis and if possible to develop biomarker predictors of this disorder.

Studies have now confirmed the clinical feasibility of first-trimester screening for early-, late-, and intermediate-onset varieties of PE using demographic, clinical, biomarker, and uterine artery Doppler information.^{12,13} Recently, the National Collaborating Center for Women’s and Children’s Health in the United Kingdom issued clinical guidelines¹⁴ for routine early prenatal screening for PE based on

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Received June 18, 2012; revised Nov. 4, 2012; accepted Nov. 8, 2012.

This study was partly supported by a grant from the Fetal Medicine Foundation, Charity Number 1037116.

The authors report no conflict of interest.

Reprints not available from the authors.

0002-9378/\$36.00

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<http://dx.doi.org/10.1016/j.ajog.2012.11.003>

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TABLE 1
Demographic and other characteristics: late-onset preeclampsia vs control group

| Parameter | Late-onset preeclampsia | Control | P value |
|---|-------------------------|-------------|---------|
| No. of cases | 30 | 59 | — |
| Maternal age, y, mean (SD) | 31.2 (6.4) | 30.8 (5.6) | .81 |
| Racial origin, n (%) | | | .02 |
| White | 14 (46.7) | 44 (74.6) | |
| Black | 14 (46.7) | 14 (23.7) | |
| Asian | 0 (0) | 1 (1.7) | |
| Mixed | 2 (6.7) | 0 (0) | |
| Nullipara, n (%) | 12 (40) | 31 (52.5) | .37 |
| Weight, kg, mean (SD) | 74.9 (15.7) | 67.7 (12.2) | .03 |
| Crown-rump length, mm, mean (SD) | 62.0 (9.1) | 62.7 (7.6) | .69 |
| Uterine pulsatility index, MoM, mean (SD) | 1.07 (0.35) | 0.98 (0.31) | .22 |

MoM, multiples of median.

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maternal demographic, historical, and clinical characteristics. It is possible that, in the future, combining clinical with biomarker predictors could further enhance screening accuracy. Our primary objective was to evaluate the use of metabolomics to identify first-trimester biomarkers of late-onset PE. Secondly, we evaluated the diagnostic accuracy of these markers for late-onset PE prediction. Finally, we evaluated the capability of metabolomics for distinguishing late- from early-onset PE.

MATERIALS AND METHODS

Study population

This study is part of an ongoing prospective study being conducted by the Fetal Medicine Foundation, London, United Kingdom, for the first-trimester prediction of important fetal and obstetric disorders. Institutional review board project #02-03-033 approval was obtained on March 14, 2003. The details of patient evaluation and study methods have been extensively described in a prior report of metabolomic prediction of early-onset PE.¹⁵ A routine population of British women was prospectively screened from March 2003 through September 2009 and they all gave written consent to participate in the study, which was approved by the King's College Hospital Research Eth-

ics Committee. Briefly, women were recruited at 11⁺⁰–13⁺⁶ weeks' gestation. Maternal characteristics and medical history were documented and first-trimester ultrasound, including crown-rump length (CRL) and uterine artery Doppler pulsatility index (PI), was measured. Data collection was planned before laboratory testing. The lower of the left and right uterine artery Doppler PI value was used for PE prediction. Maternal serum samples were also obtained and stored at –80°C for subsequent laboratory analysis. The long-term objective of the project is to develop and evaluate new markers and existing biomarkers of PE. Apart from the previously published study on early-onset PE¹⁵ these cases represent the first metabolomic analysis of PE from this study population.

A total of 30 singleton pregnancies that subsequently developed late-onset PE requiring delivery >34 weeks formed the study group and were matched with 60 unaffected controls. These cases were not previously used in any prior publication. The late-onset PE cases were selected at random from our database of available stored samples. Each case of late-onset PE was matched with 2 controls who delivered a phenotypically normal neonate with appropriate birth weight for gestational age at term and did not develop any hypertensive

disorder of pregnancy and who had blood collected within 3 days of assessment of the late-onset PE case. There was no evident source of bias in the selection of cases or controls. The definition of PE used was that proposed by the International Society for the Study of Hypertension in Pregnancy,¹⁶ namely systolic pressure ≥ 140 mm Hg or diastolic pressure ≥ 90 mm Hg on ≥ 2 occasions 4 hours apart >20 weeks of gestation, in women who were previously normotensive. Proteinuria was defined as a total of 300 mg in a 24-hour urine collection or 2 readings of at least 2⁺ proteinuria on a midstream or catheterized urine specimen in the absence of a 24-hour urine collection must also have been present in addition to the hypertension. Proteinuria must also have been present in addition to the hypertension for the diagnosis of PE. No HELLP syndrome or gestational hypertension cases were included.

Nuclear magnetic resonance (NMR) spectrometry was used for metabolite identification in the specimen samples. Sample preparation and NMR spectroscopy methods were performed as detailed previously using a 500-MHz Varian Inova NMR spectrometer (Varian Inc, Palo Alto, CA).¹⁵ Forty metabolites in each serum sample were identified and quantified in each case and control sample using commercial (NMR Suite 7.1; Chenomx Inc, Edmonton, Alberta, Canada) spectral fitting software containing an NMR spectral reference library of >200 compounds. On initial metabolomic analysis, readers were blinded to patient status.

Statistical analysis

Recommended statistical procedures for metabolomic analysis^{3,17} were followed. Log scaling was used for normalization of all metabolomic data. Principal component analysis (PCA) and partial least squares discriminant analysis (PLS-DA) were performed to identify patterns.³ PCA is an unsupervised classification technique for transforming a complex collection of data points such that the important properties of the sample can be more simply displayed along the X- and Y-axes. Two clusters on the PCA plot indicate that there

are significant metabolite differences between the normal and control groups.

PLS-DA is used to enhance the separation between the groups by summarizing the data into a few latent variables that maximize covariance between the response and the predictors.¹⁸ To minimize the possibility that the observed separation on PLS-DA is due to chance, permutation testing was performed. This involved repeated (2000 times) data sampling, with different random labeling. A significant *P* value indicates that the separation observed between groups is very unlikely to be due to chance. The MetaboAnalyst computer program was used to perform (PCA and PLS-DA) analyses.¹⁹ A variable importance in projection (VIP) plot¹⁸ is a plot ranking the metabolites based on their importance in discriminating study from control groups. Metabolites with the highest values on X- and Y-axes are the most powerful group discriminators.

In comparing the concentrations of metabolites between groups, outlier testing was performed using Dixon Q test.²⁰ The Dixon Q test is used for identification of outliers in the dataset and replaces that value with the one closest to it. Replacement of outliers helps to meet the assumption of normal distribution and equal variance between groups. Only a single value (for valine) was adjusted in this fashion, however. Kolmogorov-Smirnov and Shapiro-Wilk tests of normal distribution were performed. Metabolite concentrations in late-onset PE vs controls were compared using the 2-tailed *t* test. Mann-Whitney *U* test was used in comparing metabolite concentrations between groups that were not normally distributed. Other independent variables including fetal CRL, uterine artery Doppler, PI, and maternal age, parity, weight, ethnicity, smoking, and medical disorders were included in the genetic computing analyses along with the metabolite concentrations for PE prediction.

Genetic programming is a branch of evolutionary computing while genetic computing is a branch of genetic programming. The advantage of genetic computing lies in its ability to handle nonnormally distributed outcome measures and the large volume of data generated from “om-

TABLE 2
Serum metabolite concentrations by nuclear magnetic resonance

| Metabolite | Late-onset PE, mean (SD) (concentration in $\mu\text{mol/L}$) | Controls, mean (SD) (concentration in $\mu\text{mol/L}$) | <i>P</i> value | Fold change |
|----------------------|--|---|----------------|-------------|
| No. of cases | 30 | 59 | — | — |
| Glycerol | 800.7 (541.7) | 312 (296.8) | < .001 | 2.4 |
| 1-methylhistidine | 70.3 (40.0) | 38.9 (20.3) | < .001 | 1.7 |
| Valine | 142.5 (50.6) | 121.6 (43.3) | < .05 | 1.1 |
| Carnitine | 46.8 (24.8) | 27.8 (20.0) | .001 | 1.7 |
| Acetone | 22.1 (11.4) | 14.9 (8.5) | .003 | 1.6 |
| Trimethylamine | 6.03 (2.0) | 7.6 (3.3) | .005 | 0.88 |
| Isopropanol | 10.7 (4.6) | 7.7 (4.8) | .006 | 1.4 |
| Pyruvate | 83.1 (45.8) | 62.1 (24.1) | .006 | 1.3 |
| Hydroxyisovalerate_3 | 6.5 (3.3) | 4.7 (2.5) | .008 | 1.4 |
| Acetamide | 11.9 (7.8) | 16.1 (6.4) | .008 | 0.73 |
| Glucose | 4312.9 (1783.0) | 3362.4 (765.9) | .008 | 1.2 |
| Dimethylamine | 3.2 (1.7) | 4.4 (2.1) | .012 | 0.74 |
| Hydroxybutyrate_2 | 28.0 (14.4) | 21.2 (7.5) | .02 | 1.3 |
| Creatinine | 63.2 (16.5) | 55.1 (14.7) | .021 | 1.1 |
| Creatine | 41.5 (5.9) | 33.4 (15.6) | .024 | 1.2 |
| Citrate | 85.9 (26.9) | 74.1 (23.5) | .028 | 1.3 |
| Hydroxybutyrate_3 | 49.9 (46.7) | 29.7 (19.1) | .038 | 1.4 |
| Leucine | 114.5 (98.5) | 87.1 (61.9) | .112 | 1.2 |
| Acetate | 80.6 (101.5) | 49.1 (52.5) | .12 | 1.6 |
| Betaine | 33.3 (23.6) | 21.6 (9.4) | .14 | 1.5 |
| Glutamine | 253.1 (131.1) | 218.5 (66.9) | .182 | 1.2 |
| Ethanol | 67.7 (42.6) | 56.1 (37.2) | .19 | 1.2 |
| Ornithine | 36.8 (17.4) | 42.3 (22.5) | .24 | 0.87 |
| Acetoacetate | 18.9 (9.8) | 16.5 (9.6) | .27 | 1.1 |
| Alanine | 366.8 (204.8) | 323.8 (151.2) | .27 | 1.1 |
| Lactate | 1213.1 (564.7) | 1100.9 (689.3) | .44 | 1.1 |
| Methionine | 24.7 (7.4) | 23.6 (6.5) | .48 | 1.0 |
| Threonine | 157.2 (60.2) | 166.2 (62.5) | .5 | 0.94 |
| Propylene glycol | 11.1 (5.0) | 11.8 (4.9) | .51 | 0.93 |
| Formate | 27.0 (13.8) | 29.0 (17.8) | .6 | 1.02 |
| Tyrosine | 65.1 (23.7) | 62.3 (21.7) | .6 | 1.04 |
| Proline | 172.2 (57.7) | 165.7 (56.4) | .61 | 1.04 |
| Serine | 148.4 (103.4) | 158.6 (92.2) | .635 | 0.93 |
| Arginine | 136.3 (55.5) | 131.2 (35.9) | .65 | 1.04 |
| Asparagine | 31.3 (11.3) | 32.4 (13.7) | .71 | 0.96 |
| Phenylalanine | 78.0 (45.9) | 80.9 (45.4) | .78 | 0.96 |
| Glycine | 238.4 (129.3) | 244.0 (115.7) | .84 | 0.97 |

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(continued)

TABLE 2

Serum metabolite concentrations by nuclear magnetic resonance (continued)

| Metabolite | Late-onset PE, mean (SD) (concentration in $\mu\text{mol/L}$) | Controls, mean (SD) (concentration in $\mu\text{mol/L}$) | P value | Fold change |
|-------------|--|---|---------|-------------|
| Choline | 172.3 (341.5) | 185.7 (351.6) | .87 | 0.93 |
| Succinate | 13.2 (13.8) | 13.4 (12.9) | .9 | 0.83 |
| Malonate | 23.1 (14.3) | 23.1 (8.7) | .97 | 0.98 |
| Isobutyrate | 7.6 (2.5) | 7.6 (3.2) | 1 | 1.0 |

PE, preeclampsia.

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ics" laboratory analyses²¹ and it has previously been successfully used for metabolomic analysis.^{15,21} Genetic computing generates rules by which an optimal number of variables can be selected from a large number of exploratory variables for the optimal prediction of the outcomes of interest. The Gmax computer program, version 11.09.23 (Genetic Computing Consultants Limited, London, UK) was used for genetic computing analysis.

In addition, using a limited number of independent biochemical and demographic predictor variables, logistic regression was used to generate probability equations for the prediction of PE. Based on the regression equations generated,

individual probability for developing late-onset PE was calculated. Receiver operating characteristics curves with sensitivity plotted against false-positive rates (1-specificity) were generated along with the 95% confidence interval and/or P values for the area under the curve. For the primary analysis, power analysis indicated that a minimum of 17 cases was needed in each group to have 80% power for a 2-sided $P < .0001$.

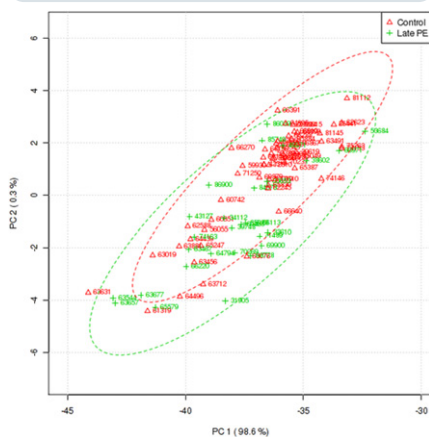
After the primary analysis, a normal group of 60 control cases previously reported for the early-onset PE metabolomic study¹⁵ derived from the same patient population was subsequently added to the current study group of 30 cases and 59 control patients to permit sufficient power to perform the regression analyses.²² Finally, we used PCA and PLS-DA analyses to determine whether late-onset PE could be differentiated from early-onset PE by directly comparing the 30 early-onset PE cases reported in a prior publication¹⁵ with the 30 late-onset PE cases in the current study.

RESULTS

Results for a total of 30 cases and 59 (of 60) controls used for the primary analysis are reported, as insufficient volume of serum was available for metabolomic analysis in one of the control samples. Table 1 compares the maternal age, weight, race, and gestational age at blood collection was determined by and represented by CRL measurements, between late-onset PE and normal cases. There was a significant difference in maternal race with a lower percentage of whites and higher percentage of blacks in the

late-onset PE group as well as greater body weight in the PE group (Table 1). In Table 2 we assessed the pairwise differences between individual metabolite concentrations for controls and late-onset PE patients. A total of 17 metabolites were present in significantly different concentrations in late-onset PE vs control groups. Fourteen of these 17 metabolite concentrations were increased in the late-onset PE group while 3 were reduced compared to normals. The P values determined via a Student *t* test prove that for 17 of the metabolites, the individual concentration differences are significant (meaningful and reproducible) at levels determined by $P < .05$. The fold changes are also presented in Table 2. Some separation and discrimination is achieved between the cases of late-onset PE and the controls from the PCA analysis of the NMR data is shown in Figure 1. The separation, however, did not appear dramatic. The PLS-DA analysis resulted in a detectable separation between late-onset PE group (in green) compared to normal cases (in red) (Figure 2). Permutation testing revealed that the observed separation of the late-onset PE from the normal group was highly unlikely to be due to chance ($P < .0005$). Figure 3 displays the VIP plot. Glycerol, carnitine, methylhistidine, and acetone appeared to be the most important metabolites for distinguishing late-onset PE from normal cases, based on VIP analysis. A heat map is shown on the right of the VIP plot. Red indicates that a particular metabolite concentration is increased in late-onset PE cases while green indicates reduced concentration compared to normal controls.

In Table 3, using genetic computing analysis of the primary dataset, ie, 30 late-onset PE cases and 59 controls, high diagnostic accuracy for late-onset PE detection was achieved. First-trimester uterine artery Doppler measurements were not significantly different between case and control groups (Table 1) and did not improve the model for late-onset PE prediction. Logistic regression-based prediction, using an expanded number of subjects, ie, 30 late-onset PE and a total of 119 normal cases, similarly showed good diagnostic accuracy for late-onset PE prediction (Table 4). The

FIGURE 1
Principal components analysis plot

Principal component analysis plot showing some separation between late-onset preeclampsia (PE) (green) and control (red) for nuclear magnetic resonance spectrometry.

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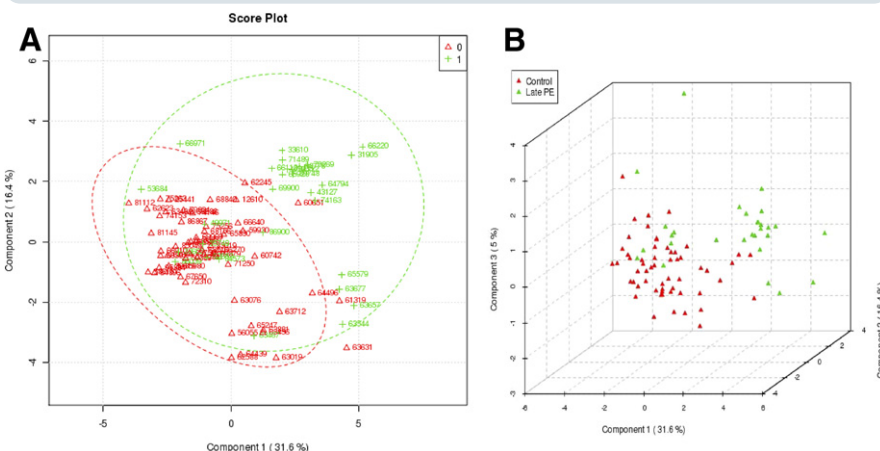
respective derived probability equations for PE are also provided in the legend of Table 4. For the regression-based analysis a limited number of predictors, glycerol and carnitine, were chosen based on the VIP plot (Figure 3) along with either maternal race or alternatively body weight. The latter 2 are known to be risk factors for late-onset PE.²³ One-methylhistidine was also combined with glycine and carnitine to form a biochemistry-only algorithm. There was no significant correlation noted between maternal weight or racial status and glycine, carnitine, or 1-methylhistidine in either the late-onset PE ($P \geq .37$) or normal ($P \geq .36$) patient group. Neither maternal weight nor race appeared to significantly enhance PE prediction over these metabolites alone (Table 4) in the regression-based analyses.

Importantly, significant discrimination between early- and late-onset PE was achieved using metabolomics (Figure 4, A). This result appears to support the current view that these are different disorders. Glycerol, acetate, trimethylamine, and succinate appeared to be the most important metabolites for distinguishing the 2 types of PE based on VIP analysis shown in Figure 4, B.

COMMENT

Using NMR-based metabolomic analysis we found 17 metabolites that were present in statistically significantly different concentrations in late-onset PE cases compared to normal controls. Using metabolites by themselves or combined with traditional maternal demographic and clinical markers, significant diagnostic accuracy for late-onset PE detection was achieved. Late-onset PE is more common than the early-onset variety and can be associated with significant morbidity,¹¹ justifying interest in its detection. We were also able to distinguish early- from late-onset PE using metabolomics. The heterogeneous etiology¹⁰ of late-onset PE has posed a particular challenge in developing predictive biomarkers. This heterogeneity is however a strength and not a limitation for metabolomic analysis. The findings are all the more interesting given the long interval between testing and disease

FIGURE 2
Partial least squares plots of metabolic data



A, Two-dimensional (2D) and **B**, 3-dimensional (3D) partial least squares discriminant analysis separations using nuclear magnetic resonance–based metabolomic measurements of late-onset preeclampsia (PE) cases (green) versus controls (red). Clear clustering and segregation of 2 patient groups indicate that significant discrimination of groups is achieved based on metabolite concentration differences. (30 early-onset PE cases/59 controls.)

*2000 permutations or resamplings were performed ($P < .0005$).

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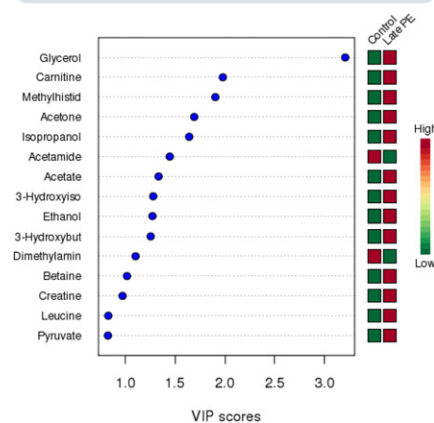
manifestation. Metabolites are known to be stable when stored at -80°C and since the cases and controls were collected at the same time and stored under identical conditions variability due to storage is unlikely to have accounted for the observed differences between groups. First-trimester prediction of PE could potentially facilitate the use of prophylactic aspirin <16 weeks of gestation.

The fact that the fold changes in metabolite concentrations seen in our study are not very large is not unexpected. Blood must be maintained in very tight homeostasis. In some cases, a 2-fold change in vital metabolite levels (eg, glucose or creatinine) represents the difference between health and disease. A 3-fold change can mean the difference between life and death. Further, late-onset PE is an extremely heterogeneous disorder,¹⁰ thus a single biomarker that definitively distinguishes (very high fold change) this disorder from normal would therefore be both biologically implausible and would obviate the need for metabolomic analysis. As stated, the advantage of metabolomic analysis for complex disorders is the ability to identify a constellation of markers that individually have modest diagnostic accu-

racy but when combined is strongly predictive of the disorder.

The diagnostic accuracy of the metabolites combined with race and/or weight

FIGURE 3
VIP plot: most discriminating metabolites in descending order of importance



Color boxes indicate whether metabolite concentration is increased (red) or decreased (green) in preeclampsia (PE) versus control. (30 early-onset PE cases/59 controls.)

VIP, variable importance in projection.

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TABLE 3
Prediction of late-onset preeclampsia based on genetic computing (primary dataset^a)

| Model | Sensitivity, % | Specificity, % | AUC | P value |
|---------------------------|----------------|----------------|-------|----------|
| Parsimonious ^b | 60 | 96.6 | 0.885 | < .00001 |
| Complex ^c | 76.7 | 100 | 0.96 | < .00001 |

AUC, area under curve.

^a 30 early-onset preeclampsia cases/59 controls; ^b Methylhistidine, glycerol, weight, race, acetoacetate; ^c Valine, weight, race, and others (pyruvate, Hydroxybutyrate_3, 1-methylhistidine, glycerol, trimethylamine, medical disorder). Other metabolites account for 5.1% of model prediction of preeclampsia.

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found in this study appears higher than that reported for widely available traditional clinical markers used by themselves. In a large prospective study of first-trimester prediction of late-onset PE, maternal risk factors and systolic, diastolic, and mean arterial pressures had sensitivities of 41.4%, 31.2%, 33.6%, and 36.7%, respectively, at a false-positive rate of 10%.²³ The concentrations of the 3 most metabolomic markers were found to be independent of maternal weight or ethnicity. We also presented evidence that metabolomic analysis can distinguish cases destined to develop early- vs late-onset PE cases.

Very little has been published on the use of metabolomics for PE prediction. The limited data available, including ours, uniformly support the value of this discovery tool.²⁴⁻²⁶ Interestingly, there was little overlap between the metabolites of significant diagnostic value in the current manuscript compared to these publications. The explanation for differences could be due to a number of factors.

We used a quantitative NMR-based platform that precisely determined the metabolite concentrations while Kenny et al²⁴ used a semiquantitative liquid chromatography (LC)-mass spectrometry (MS)-based platform. Despite some overlap, different metabolomic platforms used in the 2 studies are known to identify largely different metabolites.^{1,3} For example, NMR identifies polar compounds, while LC-MS identifies nonpolar compounds. In addition, the metabolites identified by the LC-MS study of Kenny et al²⁴ were obtained via mass matching only, whereas the metabolites identified/quantified in the current study were identified based on authentic standards and comprehensive spectral matching. Similarly, another study by Kenny et al²⁵ used an electrospray ionization MS method and the metabolites found to be significant predictors of PE were not surprisingly largely different from those reported here. A study by Odibo et al²⁶ using an LC/MS-MS platform reported accurate first-trimester prediction of PE. Again the

significant metabolites identified were primarily amino acids and largely differed from those reported here. Their manuscript focused primarily on early-onset PE cases. At this stage, the different capabilities of the metabolomic platforms is an overall positive as it allows a more comprehensive search for potentially useful markers. Ultimately, large prospective studies would be needed to identify the optimal combination of clinically useful markers for PE screening and which platform might be more advantageous clinically.

Although not a primary objective of this study, metabolomics is known to be a powerful tool for helping to elucidate the pathogenesis of complex disorders. A review of the Human Metabolomics Database¹ was performed to determine the role of some of the metabolites noted to be abnormal in late-onset PE. Two metabolites based on the VIP plot (Figure 3 and Table 2) were particularly notable for their differences between the PE and normal groups: glycerol and carnitine. Glycerol is a 3-carbon alcohol that forms the backbone of glycolipids. It is therefore important in lipid metabolism. Abnormalities of lipid metabolism are recognized to be a feature of PE and are thought to play a role in its pathogenesis.²⁷ Further, there is a well-documented relationship between maternal obesity and an increased risk of late-onset PE,²⁸ which we also found in this study. Carnitine is a quaternary ammonium compound that is responsible for the transport of lipids from the cytoplasm into the mitochondria for energy metabolism. It is made primarily in the liver and kidneys, which are both significantly affected in PE. Carnitines inhibit oxidative stress and prevent lipid peroxidation. Both of these are important pathological processes in PE. Increased carnitine production could be a response plausibly intended to counter excessive oxidative metabolism of lipids. Based on the metabolomic data found here, a picture of a central disturbance in lipid metabolism in late-onset PE emerges.

In conclusion, first-trimester metabolomic markers appeared preliminarily to

TABLE 4
Prediction of late-onset preeclampsia based on logistic regression model (expanded dataset^a)

| Model | Sensitivity, % | Specificity, % | AUC (95% CI) | P value |
|--|----------------|----------------|---------------------|---------|
| Glycerol ^b | 40 | 94.1 | 0.79 (0.692–0.888) | < .001 |
| Glycerol and weight ^c | 40 | 95 | 0.796 (0.698–0.894) | < .001 |
| Glycerol, 1-methylhistidine ^d | 56.7 | 95 | 0.783 (0.667–0.898) | < .001 |

Respective probability equations based on the regression analyses.

AUC, area under curve; CI, confidence interval.

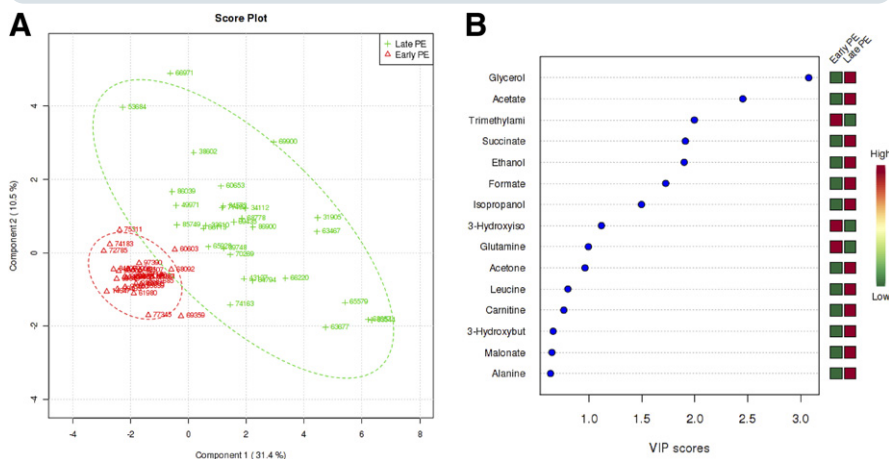
^a Sixty normal cases added from prior publication¹⁵ (total 30 late-onset preeclampsia and 119 normals); ^b Predictors considered in regression: glycerol, carnitine, and white/non-white race. Prob (preeclampsia) = 0.002*glycerol-2.60;

^c Predictors considered in regression: glycerol, carnitine, and weight. Prob (preeclampsia) = 0.002*glycerol + 0.033*weight;

^d Predictors considered in regression: glycerol, carnitine and 1-methylhistidine. Prob (preeclampsia) = 0.002*glycerol + 0.032*methylhistidine-4.04.

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FIGURE 4
PLS-DA and VIP plots: early- and late-onset PE



A, Partial least squares discriminant analysis (PLS-DA) plot showing separation between 30 early-onset (*green*) and 30 late-onset (*red*) preeclampsia (PE) cases with nuclear magnetic resonance analyses. *P* value for permutation test is .0005, which is statistically significant. **B**, Early- vs late-onset PE. Metabolites ranked by variable importance in projection (VIP).

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be useful in discriminating late-onset PE from normal controls or early-onset PE cases. Based on these promising preliminary results, the value of metabolomics for clinical prediction and diagnosis of PE should be further investigated. ■

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